

Inflammation in Asthma Pathogenesis: Role of Eosinophil, Basophils, Neutrophil and Mast Cells.

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Abstract

Asthma is a common chronic airway disease worldwide. Recent studies findings suggested that asthma is not a single disease entity, but it is a syndrome that characterised shortness of breath, intermittent attack, cough and wheezing. The disease onset commonly occurred during childhood. The natural history of asthma is variable and the disease not restricted to children, also it affects adults and both gender. Asthma pathogenesis is a complex scenario interplay of sequences of inflammation, immune responses, and airway remodelling. Thus the disease presented with different phenotypes and endotypes which are with variable treatment outcomes. The chronic inflammation is a main corner stone of asthma pathogenesis during attack and during asymptomatic course. The inflammation and immune responses are induced by the inhaled allergens and multiple host cells are involved in asthma pathogenesis. In this review we present the role of eosinophils, basophils and neutrophils in pathogenesis of asthma.

Keywords: Asthma, eosinophil, basophil, neutrophil.

1. Introduction

Asthma is a common chronic airway disease worldwide [1,2]. Asthma is not a single disease entity, but it is a syndrome that characterised by shortness of breath, intermittent attack, cough and wheezing. The disease onset commonly occurred during childhood [3]. The natural history of asthma is variable and the disease not restricted to children, also it affects adults and both gender [3]. Asthma pathogenesis is a complex scenario interplay of sequences of inflammation, immune responses, and airway remodelling. [4,5]

The chronic inflammation is a main corner stone of asthma pathogenesis during attack and during asymptomatic course [6]. The inflammation and immune responses are induced by the inhaled allergens, that include house dust mite, moulds, grasses, pollen, animal dander, and trees [7,8]

After the induction phase of the disease, there is possibility of remission or course chronicity with intermittent attacks which lead to smooth muscle hypertrophy, epithelial mucus metaplasia, and glycoprotein deposition in the sub-epithelial matrix [6]. When allergens come in contact with mucus and epithelial barriers penetrate these barriers and induce cytokines that induce subsequent events. Viral infections is a major risk factor for

asthma induction mainly in atopic individuals [9]. Chronic inflammation in asthma was initiated and driven by interplay of innate immune response, adaptive immune response and respiratory epithelium [10].

2. Asthma phenotypes and endotypes

Asthma is not a single entity and it is a heterogenic syndrome. Asthma is defined as a *"collection of several distinct diseases (endotypes) and varying phenotypes (young atopic, obese middle aged and elderly), all of which manifest with symptoms of wheezing and shortness of breath to cough and chest tightness and accompanied by variable airflow obstruction"*[11]. Table 1. Shows the endotypes and phenotypes.

Table 1. Asthma endotypes and phenotypes. [11].

| Endotype | Phenotype | Clinical features | Molecular mechanism | Biomarkers | Natural history |
|----------|-----------------|--|--|--|---|
| T2 high | Atopic | Early onset, well defined, steroid sensitive | Allergic sensitisation | Blood/sputum eosinophils count, serum specific allergen IgE, high FeNO, high total IgE | Identifiable and treatable, preserved lung function |
| | Late onset | ± concomitant CRSwNP, steroid refractory | Staphylococcus aureus enterotoxin | Blood/sputum eosinophils count, high FeNO, | Severe from onset, more frequent exacerbation |
| | AERD | Adult onset | Dysregulated arachidonic acid metabolism | Blood/sputum eosinophils count, urinary LTE4 | Severe from onset, more frequent exacerbation |
| Non-T2 | Non-atopic | Adult onset, paucigranulocytic or neutrophilic | NLRP3/IL-1 β , altered micro-RNA expression, Th17 | Induced sputum neutrophil count, MMP-9 in Bal | Variable course and lung function |
| | Smokers | Older adult | Oxidative stress, mixed Th2 high Th2 low | Induced sputum neutrophil count | More frequent exacerbation, low lung function |
| | Obesity related | Female sex | Oxidative stress, neutrophils, increased innate immune activation, | Serum IL-6 | Severe symptoms, preserved lung function |
| | Elderly | >50 to > 65 years at onset | Immunosenescence, Th1/Th17 inflammation | Induced sputum neutrophils count | Steroid resistant |

CRSwNP= Chronic rhinosinusitis with nasal polyp

FeNO= Fractional excretion of Nitric Oxide

3. Eosinophils role in induction of inflammation in asthma

The role of eosinophils in pathogenesis of asthma was reviewed in last decade [12-15]. The measurement of the inflammation of airway that was induced by eosinophils can be achieved invasively by bronchoscopic sampling or non-invasive sputum analysis [13]. Sputum and bronchoalveolar lavage examined for eosinophils counts, cytology, FeNO, eosinophil cationic protein, eosinophil-derived neurotoxin, eosinophil peroxidase, and major

basic protein. Eosinophil recruitment from blood to tissue required eosinophils activation [16-18]. Allergic diseases including asthma were associated down-regulation or up-regulation of surface eosinophils proteins and Fc receptors or integrins activation [16-19]. Eosinophils surface proteins such as CD44, CD45RO, CD48, CD137, CD89, CD16, IL-2R α , IL-17RA, CD25, IL-17RB, regulated in asthma and involved in disease pathogenesis [16, 20,21].

The eosinophils surface proteins that serve as a target for therapy in asthmatic patients include activated β 2 integrin, CD162, CD125, CD25, CD18, CD11a, and CD11b [16,21]. However, eosinophils express several inhibitory receptors [22].

The mediators that were released by eosinophils had the capacity for induction of airway hyperresponsiveness [15]. Human eosinophils major basic proteins and eosinophils peroxidase induced airway hyperresponsiveness (AHR) in animal models [23,24]. However, eosinophils cationic proteins and eosinophils-derived neurotoxins did not [23]. The mechanism by which AHR was induced by bradykinin production [24]. In addition, major basic proteins cause histamine release by basophils and mast cells [25,26].

Eosinophils count in sputum and bronchoalveolar lavage fluid was lower in none asthmatic as compared to asthmatic subjects [27]. Th2 cytokines expression like IL-5 as an indicator of eosinophilic inflammation increased in bronchoalveolar lavage fluid from asthma patients [28]. Eosinophilia of blood correlated with asthma exacerbation frequency and severity [27,29], however, there was none eosinophils asthma phenotype and presence of other causes for peripheral eosinophilia [30,31]. Several cytokines such as IL-13 produced by eosinophils and thus IL-13 lead to AHR and induce mucus secretion [24]. Th2 cells and ILC2s also produced IL-13. In addition, eosinophils produced leukotrienes which induce AHR [32].

Eosinophils development from CD34⁺ hematopoietic progenitor cells were promoted by IL-5, IL-3 and granulocyte-macrophage colony stimulating factor, however, only IL-5 was specific for eosinophils development [12]. Proinflammatory mediators are produced by eosinophils which included eosinophils cationic protein, cytokines and newly synthesised eicosanoids [33-35]. Previous studies [12-15] suggested that eosinophils play a role in asthma pathogenesis through release of mediators, cytokines and chemokines such as MBP, ECP, EDN, EPO, galectin-10, LTC₄, PEG2, platelet-activating factor, thromboxane, TGF- β , IL-3, IL-4, IL-5, IL-8, IL-10, IL-12, IL-13, IL-16, IL-18, TNF- α , CCL5 and CCL11. Airway remodelling was a potential change that was caused by eosinophils inflammation induction and release of fibrogenic mediators and multiple growth factors such as transforming growth factor (TGF- β), matrix metalloproteinase-9, tissue inhibitor of metalloproteinase-1, vascular endothelial growth factor, basic fibroblast growth factor, angiogenin, MBP, ECP, IL-17, IL-13, heparin-binding epidermal growth factor, nerve growth factor, cystinyl leukotrienes, and stem cell factor [36-55]. In asthma exacerbation, eosinophilia of the airway was the early feature [56]. In addition, eosinophils released cytokines were responsible for induction of various immunomodulation in asthma patients [26,57-70]. Fig1. Illustrate the role of eosinophils in asthma pathogenesis.

4. Basophils role in induction of inflammation in asthma

Recent studies indicated that basophils play a potential role in induction of allergic inflammation in both IgE dependent and none-dependent. [71]. Basophils migrated to the site of allergic inflammation and secreted chemokines, proteases and cytokines [12]. The mediators that were produced by basophils divided in to cytokines/chemokines, preformed mediators, and newly synthesised lipid mediators [72]. In allergic inflammation basophils induced its effect through histamine that is stores in granules, rapid production of LTC₄, LTD₄, and LTE₄ which cause bronchoconstriction and increase vascular permeability [12,14].

Activated basophils expressed cytokines such as GM-CSF, IL-13 and IL-4, but IL-4 was secreted by activated basophils in high concentration and rapid response [12]. In animal model, basophils production of none mediated IgE IL-4 was the early differentiation of Th2 [73]. IgE synthesis amplification under the control of expression of IL-13, IL-4 and CD154 by basophils [12]. A novel mediator, granzyme B (protease) was secreted in asthma following challenge with inhalation allergens [74]. The predominant source of IL-4 in allergen activated polymorphonuclear cells and in mouse models [12]. Post-mortem studies on patients die due to asthma show increased number of basophils in lung tissue [73,75-77]. Studies in animal models suggested that basophils play direct role as antigen presenting cell that lead to Th2 responses induction, IL-4 and MHC class II molecule expression [78-80]. CD63 and CD203C are the well-described human basophils activation markers [81-83]. Similar up-regulation exhibited by CD107a, CD164 and CD13 [84]. Identification of basophils can be achieved by expression of CCR3, CD123, or CRTH2 cell surface markers [82,83]. Additionally, basogranulin was a the specific basophils marker [85] and secreted after both non- IgE and IgE- mediated stimuli [86]. Basophils identification in tissue can be done using immunohistochemical technique to detect basogranulin as specific marker [87].

5. Neutrophils role in induction of inflammation in asthma

Neutrophilic asthma is one of the refractory asthma phenotype, which characterised by course severity, fixed obstruction of the airway, poor response to treatment and frequent exacerbation [88-91]. Neutrophilic asthma was not fully understood phenotype, however, this phenotype was complex and form about 30-50% of symptomatic asthma [92]. Obesity, gastrointestinal reflux disease, respiratory infections, and obstructive sleep apnoea were associated with neutrophilic asthma [93].

Better outcomes of severe asthma treatment were achieved recently by the phenotypes characterization which contribute to personalized therapy of asthma through the development of novel biologics [88,92,94,95,96,97]. Th2 driven eosinophilic asthma in about 50% of asthma cases, while the remaining half were none eosinophils asthma phenotypes and were subdivided into paucigranulocyte and neutrophilic subtypes [94,95,98].

Phenotype of neutrophilic asthma was less well defined [99,100], while the eosinophils asthma phenotype was well defined [96,98-105]. Neutrophilic asthma pathophysiology was complex. High neutrophils count in sputum from neutrophilic asthma in 40-76% of sputum cells [99,100,106] and less sputum eosinophils count [95,104]. Presence of high number of neutrophils in sputum was associated with persistant asthma severity [88,95,97,106,107], low FEV1 [108] and fixed airway obstruction [106,108,109]. Exacerbation was more frequent in neutrophilic asthma phenotype, but the severity was less than that in eosinophilic phenotype [110-113].

Neutrophilic asthma was characterised by nocturnal worsening which associated with high number of BAL granulocytes [114] and this guide the treatment of cases [115]. Neutrophilic asthma was with poor prognosis, worse quality of life, none responsive to high dose of inhaled corticosteroids and newly developed biologic therapy [88-90,97,102,108,116-122].

To date, there was no specific biomarkers for neutrophilic asthma diagnosis, however, the phenotype was adult onset, mainly not atopic, and with bronchoprovocation test weak responsiveness to methacholine [103,108,123-125]. Innate immune response alteration and Th17 cells activation drive the neutrophilic asthma [126-128]. IL-17A and IL-17F were play a potential role in neutrophilic asthma pathogenesis and neutrophilic inflammation. However, other cytokines and chemochines such as TGF- β , TNF- α , IL-1 β , IL-23, IL-8 and IL-6 which act with IL-17A in induction of neutrophilic inflammation in severe cases [129-132].

Pathogens elimination from the airway was done by neutrophils. But persistent neutrophilia and protease secretion will lead to airway injury, hypersecretion of mucus, and airway obstruction and remodelling [133]. Neutrophils recruitment to lung tissue in asthma patients and induce neutrophilic inflammation include the following cytokines, lipids, chemokines and complements: IL-1 α , IL-1 β , IL-1Ra, IL-6, IL-10, IL-17, IL-23, INF- γ , TNF- α , LTB₄, Lipoxin A₄, Resolvin D1/E1, PGD₂, CXCL1, CXCL5, CXCL6, CXCL8, C5a, and FMLP [134-163]. However, IL-1Ra, IL-10, Lipoxin A₄, Resolvin D1/E1, and PGD₂ were decreased while others were increased. The most potent neutrophils chemoattractant in the lung was CXCL8 [164] and was increased in asthmatic nasal secretions and sputum [165,166]. Asthmatic patients neutrophils express the high affinity IgE receptor (Fc ϵ R1) which induct release of CXCL8 from neutrophils [167]. It was suggested that Th17, Th1 and neutrophils form a network communication in airway and cause severe attack that refractory to corticosteroids [133].

In asthma patients, neutrophils form the first line defense mechanism against pulmonary infection [168], however, it release mediators that attract macrophages/monocytes to infection site [169]. Additional side effects were exerted by neutrophils and cause mucus hypersecretion, and airway obstruction, airway smooth muscle responsiveness increase, and airway remodelling [170-172]. Studies in children and adults with asthma not confirmed the association between neutrophilic inflammation and airway remodelling [173,174]. Persistence of neutrophils in airway of asthma patients may be contributed to that corticosteroids inhibition of neutrophils apoptosis [175-178], release of ATP from dying cells [179-181], LTB₄ inflammatory mediators [182], and pathogen-induced mechanisms [183].

6. Mast cells role in induction of inflammation in asthma

Mast cells activation as a response to IgE antigen binding receptor with Th2 cell activation initiate the chronic inflammation in asthma [184]. Mast cells contain high affinity IgE receptors and localised in connective tissue and mucosal membrane and blood vessels [185]. Mast cells play a potential key role in induction of inflammation and modulation of production of mediators and activated by cytokines, viral and bacterial antigens, hormones and growth factors [186]. Local production of monocytes/mast cells (MCP-1 and RANTES) chemotactic factors lead to accumulation of mast cells in inflamed tissues [187].

Mast cells with two phenotypes, the mucosal mast cells and tissue mast cells [188]. Tissue mast cells contain chymase and tryptase, while mucosal mast cells contain tryptase only, additionally, both phenotypes were differ in their number, secretory granules type and stimuli responsiveness [188]. Mucosal mast cells contain more chondroitin, while tissue mast cells contain heparin and responded to neuropeptides while mucosal mast cells not respond [188].

Mast cells play a role in asthma pathophysiology [189], and mast cells express many receptors such as CD117, Fc ϵ R1, CD32a, CD64, PGE₂ receptors, C3a and C5a receptors, adenosine receptor, β 2 adrenergic receptor, IL-10R, IL-9R, IL-5R, IL-4R, IL-3R, IFN- γ R, GM-CSFR, CXCR4, CXCR2, CCR5, CCR3, Toll-like receptor, and nerve growth factor receptor [190-192]. Mast cells release or generate PGD₂, platelets activating factor, histamine, leukotrienes, and others as a immediate inflammation responses [193]. The above mediators induce spasmogenic and vasoactive action in asthmatic patients.

Proinflammatory cytokines such as IL-8, IL-6, IL-1 and TNF- α which are innate immunity important factors and mast cells the only cells that release preformed TNF- α [194]. Theoharides et al [195] reported that asthma worsen by stress, a process that mediated by mast cells activation that infiltrate bronchial smooth muscle and cytokines secretion. IL-9 target mast cells and expanded the population of mast cells [196]. Independent of IgE, mast cells activated by IL-18 and produce histamine and Th2 cytokines [197]. Several

inflammatory cells were detected in bronchial biopsy and BAL including mast cells [198]. Mast cells synthesise IL-17 in response to signals through TLR9, TLR7, TLR4, TLR3, TLR2, and TLR1 [199]. IL-33 augmented the release of vascular endothelia growth by mast cells in response to substance P [200]. Mast cells participation in asthma development is through surface high affinity receptors for IgE cross-linking which lead to production of proinflammatory and vasoactive materials [184].

Mast cells released mediators that were classified into cytokines/ chemokines, newly synthesized lipid mediators, and preformed mediators [12], however, this categorization not absolute as that TNF- α included in newly synthesized and preformed mediators. The preformed mediators include tryptase, chymase, histamine, proteoglycans (heparin and chondroitin sulphates) carboxypeptidase A and their storage was in cytoplasmic granules [12]. These mediators play a role in the pathophysiology of asthma [14].

KIT (CD117) and Fc ϵ R1 activated mast cells and contribute to rapid synthesis of eicosanoid mediators from arachidonic acid that was stored in endogenous membrane and subsequent synthesis of cysteinyl leukotrienes which induce bronchoconstriction, attract eosinophils, mucus production induction and initiate vascular permeability. PGD₂ as newly synthesized mediators induce basophils and eosinophils attraction and bronchoconstriction [12]. Additionally, TNF- α stored and produced by mast cell and cause increase in hyperresponsiveness and upregulation of epithelial and endothelial adhesion molecules. Also mast cells release other cytokines (such as GM-CSF, IL-3, IL-5, IL-13, IL-10, IL-6) and chemokines [CCL3, CXCL8 (IL-8)] [190-192].

Based on animal studies it was suggested that mast cells to play function in adaptive immunity, innate immunity, and homeostasis [12].

Allergic diseases (asthma, allergic rhinitis and anaphylaxis) pathogenesis central action was the activation of mast cells through Fc ϵ R1 [12]. Atopic asthma specificity is the Fc ϵ R1 expression robust upregulation on nasal and alveolar mast cells and the changes of alveolar mast cells skewed toward Th2 profile that was correlated with clinical outcome [201,202].

References

1. Papi A, Brightling C, Pedersen SE, Reddel HK. Asthma. *Lancet* 2018;391(10122): 783-800.
2. Vos T, Flaxman AD, Naghavi M, et al. Years lived with disability(YLDs) for 1160 sequelae of 289 diseases and injuries 1990-2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet* 2012;380(9859):2163-2196.
3. Alsamarai AGM, Salih MA, Alobaidy AH, Alwan AM, Abdulaziz ZH. Risk factors for asthma in Iraqi children. *Jour Rural Tropical Public Health* 2009; 8:45-52.
4. Alobaidi AHA, Alsamarai AGM. Effect of acetaminophen and NAC on bronchial markers in asthma. *Middle East J Fam Med* 2007;5:8-14.
5. Alobaidi AHA, Alsamarai AGM. Evaluation of simvastatin anti-inflammatory and antioxidant effect in asthma *J Bahrain Med Society* 2008;20(2):103-108.
6. Locksley RM. Asthma and allergic inflammation. *Cell* 2010;140:777-783.
7. Alzakar RH, Alsamarai AGM. Efficacy of immunotherapy for treatment of allergic asthma in children. *Allergy Asthma Proc* 2010;31:324-330.
8. Alsamarai AGM, Alobaidi AH, Alrefaie SM, Alwan AM. House dust mite immunotherapy in Iraqi patients with allergic rhinitis and asthma. *Pharmacotherapy*. Chapter 7, pp.141-154, 2012. ISBN 979-953-305-316-2. In Tech- Open Access Publisher.
9. Gao H, Ying S, Dai Y. Pathological Roles of Neutrophil-Mediated Inflammation in Asthma and Its Potential for Therapy as a Target. *J Immunol Res*.

- 2017;2017:3743048.
10. Ishmael FT. The inflammatory response in the pathogenesis of asthma. *JAOA Supplement* 7, 2011;111(11):S11-S17.
11. Kuruvilla ME, Lee FE, Lee GB. Understanding Asthma Phenotypes, Endotypes, and Mechanisms of Disease. *Clin Rev Allergy Immunol*. 2019 Apr;56(2):219-233.
12. Stone KD, Prussin C, Metcalfe DD. IgE, mast cells, basophils, and eosinophils. *J Allergy Clin Immunol*. 2010 Feb;125(2 Suppl 2):S73-80.
13. George L, Brightling CE. Eosinophilic airway inflammation: role in asthma and chronic obstructive pulmonary disease. *Ther Adv Chronic Dis*. 2016;7(1):34-51.
14. Metcalfe DD, Pawankar R, Ackerman SJ, Akin C, Clayton F, Falcone FH, et al. Biomarkers of the involvement of mast cells, basophils and eosinophils in asthma and allergic diseases. *World Allergy Organ J*. 2016;9:7.
15. McBrien CN, Menzies-Gow A. The Biology of Eosinophils and Their Role in Asthma. *Front Med (Lausanne)*. 2017 Jun 30;4:93.
16. Johansson MW. Activation states of blood eosinophils in asthma. *Clin Exp Allergy*. 2014;44(4):482–98.
17. Johansson MW, Mosher DF. Integrin activation states and eosinophil recruitment in asthma. *Front Pharmacol*. 2013;4:33.
18. Rosenberg HF, Dyer KD, Foster PS. Eosinophils: changing perspectives in health and disease. *Nat Rev Immunol*. 2013;13(1):9–22.
19. Driss V, Legrand F, Capron M. Eosinophils receptor profile. In: Lee J, Rosenberg HF, editors. *Eosinophils in health and disease*. 1st ed. Waltham: Elsevier/Academic Press; 2013. p. 30–8.
20. Heinisch IV, Bizer C, Volgger W, Simon HU. Functional CD137 receptors are expressed by eosinophils from patients with IgE-mediated allergic responses but not by eosinophils from patients with non-IgE-mediated eosinophilic disorders. *J Allergy Clin Immunol*. 2001;108(1):21–8.
21. Simon HU, Plotz S, Simon D, Seitzer U, Braathen LR, Menz G, et al. Interleukin-2 primes eosinophil degranulation in hypereosinophilia and Wells' syndrome. *Eur J Immunol*. 2003;33(4):834–9.
22. Munitz A, Levi-Schaffer F. inhibitory receptors on eosinophils: A direct hit tp a possible Achilles heel? *J Allergy Clin Immunol* 2007;119:1382-7.
23. Gundel RH, Letts LG, Gleich GJ. Human eosinophil major basic protein induces airway constriction and airway hyperresponsiveness in primates. *J Clin Invest* 1991; 87(4):1470–3.
24. Coyle AJ, Ackerman SJ, Burch R, Proud D, Irvin CG. Human eosinophil granule major basic protein and synthetic polycations induce airway hyperresponsiveness in vivo dependent on bradykinin generation. *J Clin Invest* 1995; 95(4):1735–40.
25. Piliponsky AM, Gleich GJ, Nagler A, Bar I, Levi-Schaffer F. Non-IgE dependent activation of human lung- and cord blood-derived mast cells is induced by eosinophil major basic protein and modulated by the membrane form of stem cell factor. *Blood* 2003;101(5):1898–904.
26. Ben-Zimra M, Bachelet I, Seaf M, Gleich GJ, Levi-Schaffer F. Eosinophil major basic protein activates human cord blood mast cells primed with fibroblast membranes by integrin-beta1. *Allergy* 2013;68(10):1259–68.
27. Bousquet J, Chané P, Lacoste JY, Barnéon G, Ghavanian N, Enander I, et al. Eosinophilic inflammation in asthma. *N Engl J Med* 1990;323(15):1033–9.
28. Sur S, Gleich GJ, Swanson MC, Bartemes KR, Broide DH. Eosinophilic inflammation is associated with elevation of interleukin-5 in the airways of patients

- with spontaneous symptomatic asthma. *J Allergy Clin Immunol* 1995;96(5 Pt 1):661–8.
29. Price DB, Rigazio A, Campbell JD, Bleecker ER, Corrigan CJ, Thomas M, et al. Blood eosinophil count and prospective annual asthma disease burden: a UK cohort study. *Lancet Respir Med* 2015;3(11):849–58.
 30. Mejia R, Nutman TB. Evaluation and differential diagnosis of marked, persistent eosinophilia. *Semin Hematol* 2012;49(2):149–59.
 31. Simon HU, Rothenberg ME, Bochner BS, Weller PF, Wardlaw AJ, Wechsler ME, et al. Refining the definition of hypereosinophilic syndrome. *J Allergy Clin Immunol* 2010;126(1):45–9.
 32. Hallstrand TS, Henderson WR Jr. An update on the role of leukotrienes in asthma. *Curr Opin Allergy Clin Immunol* 2010;10(1):60–6.
 33. Blanchard C, Rothenberg ME. Biology of the eosinophil. *Adv Immunol* 2009;101:81–121.
 34. Hogan SP, Rosenberg HF, Moqbel R, Phipps S, Foster PS, Lacy P, et al. Eosinophils: biological properties and role in health and disease. *Clin Exp Allergy* 2008;38:709–50.
 35. Moqbel R, Lacy P, Adamko DJ, Odemuyiwa SO. Biology of eosinophils. In: Adkinson, N.; Bochner, BS.; Busse, WW.; Holgate, ST.; Lemanske, RF.; Simons, FER., editors. *Middleton's Allergy: Principles and Practice*. 7th ed. Mosby Elsevier; 2009.
 36. Phipps S, Ying S, Wangoo A, Ong YE, Levi-Schaffer F, Kay AB. The relationship between allergen-induced tissue eosinophilia and markers of repair and remodeling in human atopic skin. *J Immunol* 2002;169(8):4604–12.
 37. Vignola AM, Chanez P, Chiappara G, Merendino A, Pace E, Rizzo A, et al. Transforming growth factor-beta expression in mucosal biopsies in asthma and chronic bronchitis. *Am J Respir Crit Care Med* 1997;156(2 Pt 1):591–9.
 38. Goldsmith AM, Bentley JK, Zhou L, Jia Y, Bitar KN, Fingar DC, et al. Transforming growth factor-beta induces airway smooth muscle hypertrophy. *Am J Respir Cell Mol Biol* 2006;34(2):247–54.
 39. McMillan SJ, Xanthou G, Lloyd CM. Manipulation of allergen-induced airway remodeling by treatment with anti-TGF-beta antibody: effect on the Smad signaling pathway. *J Immunol* 2005;174(9):5774–80.
 40. Vignola AM, Riccobono L, Mirabella A, Profita M, Chanez P, Bellia V, et al. Sputum metalloproteinase-9/tissue inhibitor of metalloproteinase-1 ratio correlates with airflow obstruction in asthma and chronic bronchitis. *Am J Respir Crit Care Med* 1998;158(6):1945–50.
 41. Johnson C, Sung HJ, Lessner SM, Fini ME, Galis ZS. Matrix metalloproteinase-9 is required for adequate angiogenic revascularization of ischemic tissues: potential role in capillary branching. *Circ Res* 2004;94(2):262–8.
 42. Hoshino M, Takahashi M, Aoike N. Expression of vascular endothelial growth factor, basic fibroblast growth factor, and angiogenin immunoreactivity in asthmatic airways and its relationship to angiogenesis. *J Allergy Clin Immunol* 2001;107(2):295–301.
 43. Thompson HG, Mih JD, Krasieva TB, Tromberg BJ, George SC. Epithelial-derived TGF-beta2 modulates basal and wound-healing subepithelial matrix homeostasis. *Am J Physiol Lung Cell Mol Physiol* 2006;291(6):L1277–85.
 44. Zagai U, Lundahl J, Klominek J, Venge P, Sköld CM. Eosinophil cationic protein

- stimulates migration of human lung fibroblasts in vitro. *Scand J Immunol* 2009; 69(4):381–6.
45. Hernnäs J, Särnstrand B, Lindroth P, Peterson CG, Venge P, Malmström A. Eosinophil cationic protein alters proteoglycan metabolism in human lung fibroblast cultures. *Eur J Cell Biol* 1992;59(2):352–63.
 46. Molet S, Hamid Q, Davoine F, Nutku E, Taha R, Pagé N, et al. IL-17 is increased in asthmatic airways and induces human bronchial fibroblasts to produce cytokines. *J Allergy Clin Immunol* 2001;108(3):430–8.
 47. Lu S, Li H, Gao R, Gao X, Xu F, Wang Q, et al. IL-17A, but not IL-17F, is indispensable for airway vascular remodeling induced by exaggerated Th17 cell responses in prolonged ovalbumin-challenged mice. *J Immunol* 2015;194(8):3557–66.
 48. Zhu Z, Homer RJ, Wang Z, Chen Q, Geba GP, Wang J, et al. Pulmonary expression of interleukin-13 causes inflammation, mucus hypersecretion, subepithelial fibrosis, physiologic abnormalities, and eotaxin production. *J Clin Invest* 1999;103(6):779–88.
 49. Malavia NK, Mih JD, Raub CB, Dinh BT, George SC. IL-13 induces a bronchial epithelial phenotype that is profibrotic. *Respir Res* 2008;9:27.
 50. Wang Q, Li H, Yao Y, Lu G, Wang Y, Xia D, et al. HB-EGF-promoted airway smooth muscle cells and their progenitor migration contribute to airway smooth muscle remodeling in asthmatic mouse. *J Immunol* 2016;196(5):2361–7.
 51. Frossard N, Freund V, Advenier C. Nerve growth factor and its receptors in asthma and inflammation. *Eur J Pharmacol* 2004;500(1–3):453–65.
 52. Huang LW, Sun G, Wang DL, Kong LF. Inhibition of nerve growth factor/tyrosine kinase receptor A signaling ameliorates airway remodeling in chronic allergic airway inflammation. *Eur Rev Med Pharmacol Sci* 2015;19(12):2261–8.
 53. Henderson WR Jr, Chiang GK, Tien YT, Chi EY. Reversal of allergen-induced airway remodeling by CysLT1 receptor blockade. *Am J Respir Crit Care Med* 2006;173(7):718–28.
 54. Kampf C, Relova AJ, Sandler S, Roomans GM. Effects of TNF-alpha, IFN-gamma and IL-beta on normal human bronchial epithelial cells. *Eur Respir J* 1999; 14(1):84–91.
 55. Sullivan DE, Ferris M, Pociask D, Brody AR. Tumor necrosis factor-alpha induces transforming growth factor-beta1 expression in lung fibroblasts through the extracellular signal-regulated kinase pathway. *Am J Respir Cell Mol Biol* 2005; 32(4):342–9.
 56. Pizzichini MM, Pizzichini E, Clelland L, Efthimiadis A, Pavord I, Dolovich J, et al. Prednisone-dependent asthma: inflammatory indices in induced sputum. *Eur Respir J* 1999;13(1):15–21.
 57. Rosenberg HF, Dyer KD, Foster PS. Eosinophils: changing perspectives in health and disease. *Nat Rev Immunol* 2013;13(1):9–22.
 58. Shenoy NG, Gleich GJ, Thomas LL. Eosinophil major basic protein stimulates neutrophil superoxide production by a class IA phosphoinositide 3-kinase and protein kinase C-zeta-dependent pathway. *J Immunol* 2003;171(7):3734–41.
 59. Kannan Y, Ushio H, Koyama H, Okada M, Oikawa M, Yoshihara T, et al. 2.5S nerve growth factor enhances survival, phagocytosis, and superoxide production of murine neutrophils. *Blood* 1991;77(6):1320–5.
 60. Kawamoto K, Okada T, Kannan Y, Ushio H, Matsumoto M, Matsuda H. Nerve growth factor prevents apoptosis of rat peritoneal mast cells through the trk proto-

- oncogene receptor. *Blood* 1995;86(12):4638–44.
61. Yang D, Chen Q, Rosenberg HF, Rybak SM, Newton DL, Wang ZY, et al. Human ribonuclease A superfamily members, eosinophil-derived neurotoxin and pancreatic ribonuclease, induce dendritic cell maturation and activation. *J Immunol* 2004;173(10):6134–42.
 62. Akuthota P, Wang H, Weller PF. Eosinophils as antigen-presenting cells in allergic upper airway disease. *Curr Opin Allergy Clin Immunol* 2010;10(1):14–9.
 63. Shi HZ, Humbles A, Gerard C, Jin Z, Weller PF. Lymph node trafficking and antigen presentation by endobronchial eosinophils. *J Clin Invest* 2000;105(7):945–53.
 64. Cocks BG, de Waal Malefyt R, Galizzi JP, de Vries JE, Aversa G. IL-13 induces proliferation and differentiation of human B cells activated by the CD40 ligand. *Int Immunol* 1993;5(6):657–63.
 65. Geha RS, Jabara HH, Brodeur SR. The regulation of immunoglobulin E class switch recombination. *Nat Rev Immunol* 2003;3(9):721–32.
 66. Chu VT, Fröhlich A, Steinhäuser G, Scheel T, Roch T, Fillatreau S, et al. Eosinophils are required for the maintenance of plasma cells in the bone marrow. *Nat Immunol* 2011;12(2):151–9.
 67. Jourdan M, Cren M, Robert N, Bolloré K, Fest T, Duperray C, et al. IL-6 supports the generation of human long-lived plasma cells in combination with either APRIL or stromal cell-soluble factors. *Leukemia* 2014;28(8):1647–56.
 68. Liu LY, Bates ME, Jarjour NN, Busse WW, Bertics PJ, Kelly EA. Generation of Th1 and Th2 chemokines by human eosinophils: evidence for a critical role of TNF- α . *J Immunol* 2007;179(7):4840–8.
 69. Odemuyiwa SO, Ghahary A, Li Y, Puttagunta L, Lee JE, MusatMarcu S, et al. Cutting edge: human eosinophils regulate T cell subset selection through indoleamine 2,3-dioxygenase. *J Immunol* 2004;173(10):5909–13.
 70. Xu H, Zhang GX, Ciric B, Rostami A. IDO: a double-edged sword for T(H)1/T(H)2 regulation. *Immunol Lett* 2008;121(1):1–6.
 71. Miyake K, Karasuyama H. Emerging roles of basophils in allergic inflammation. *Allergy International* 2017;66:382–391.
 72. MacGlashan, D. Biochemical events in basophil/mast cell activation and mediator release. In: Adkinson, N.; Bochner, BS.; Busse, WW.; Holgate, ST.; Lemanske, RF.; Simons, FER., editors. *Middleton's Allergy: Principles and Practice*. 7th ed. Mosby Elsevier; 2009.
 73. Sullivan BM, Locksley RM. Basophils: a nonredundant contributor to host immunity. *Immunity* 2009;30:12–20.
 74. Tschoep CM, Spiegl N, Didichenko S, Lutmann W, Julius P, Virchow JC, et al. Granzyme B, a novel mediator of allergic inflammation: its induction and release in blood basophils and human asthma. *Blood* 2006;108:2290–9.
 75. Karasuyama H, Mukai K, Tsujimura Y, Obata K. Newly discovered roles for basophils: a neglected minority gains new respect. *Nat Rev Immunol* 2009;9:9–13.
 76. Min B. Basophils: what they 'can do' versus what they 'actually do'. *Nat Immunol* 2008;9:1333–9.
 77. Schroeder, J. Biology of basophils. In: Adkinson, N.; Bochner, BS.; Busse, WW.; Holgate, ST.; Lemanske, RF.; Simons, FER., editors. *Middleton's Allergy: Principles and Practice*. 7th ed. Mosby Elsevier; 2009.
 78. Perrigoue JG, Saenz SA, Siracusa MC, Allenspach EJ, Taylor BC, Giacomini PR,

- et al. MHC class II-dependent basophil-CD4⁺ T cell interactions promote T(H)2 cytokine-dependent immunity. *Nat Immunol* 2009;10:697–705.
79. Sokol CL, Chu NQ, Yu S, Nish SA, Laufer TM, Medzhitov R. Basophils function as antigenpresenting cells for an allergen-induced T helper type 2 response. *Nat Immunol* 2009;10:713–20.
 80. Yoshimoto T, Yasuda K, Tanaka H, Nakahira M, Imai Y, Fujimori Y, et al. Basophils contribute to T(H)2-IgE responses in vivo via IL-4 production and presentation of peptide-MHC class II complexes to CD4⁺ T cells. *Nat Immunol* 2009;10:706–12.
 81. Uyttebroek AP, Sabato V, Faber MA, Cop N, Bridts CH, Lapeere H, et al. Basophil activation tests: time for a reconsideration. *Expert Rev Clin Immunol*. 2014;10(10):1325–35.
 82. Sturm GJ, Kranzelbinder B, Sturm EM, Heinemann A, Groselj-Strele A, Aberer W. The basophil activation test in the diagnosis of allergy: technical issues and critical factors. *Allergy*. 2009;64(9):1319–26.
 83. McGowan EC, Saini S. Update on the performance and application of basophil activation tests. *Curr Allergy Asthma Rep*. 2013;13(1):101–9.
 84. Hennersdorf F, Florian S, Jakob A, Baumgartner K, Sonneck K, Nordheim A, et al. Identification of CD13, CD107a, and CD164 as novel basophilactivation markers and dissection of two response patterns in time kinetics of IgE-dependent upregulation. *Cell Res*. 2005;15(5):325–35.
 85. McEuen AR, Calafat J, Compton SJ, Easom NJ, Buckley MG, Knol EF, et al. Mass, charge, and subcellular localization of a unique secretory product identified by the basophil-specific antibody BB1. *J Allergy Clin Immunol*. 2001;107(5):842–8.
 86. Mochizuki A, McEuen AR, Buckley MG, Walls AF. The release of basogranulin in response to IgE-dependent and IgE-independent stimuli: validity of basogranulin measurement as an indicator of basophil activation. *J Allergy Clin Immunol*. 2003;112(1):102–8.
 87. Agis H, Krauth MT, Bohm A, Mosberger I, Mullauer L, Simonitsch-Klupp I, et al. Identification of basogranulin (BB1) as a novel immunohistochemical marker of basophils in normal bone marrow and patients with myeloproliferative disorders. *Am J Clin Pathol*. 2006;125(2):273–81.
 88. Moore WC, Bleecker ER, Curran-Everett D, Erzurum SC, Ameredes BT, Bacharier L, et al. National Heart, Lung, Blood Institute's Severe Asthma Research Program. Characterization of the severe asthma phenotype by the National Heart, Lung, and Blood Institute's Severe Asthma Research Program. *J Allergy Clin Immunol*. 2007 Feb;119(2):405-13.
 89. Chanez P, Wenzel SE, Anderson GP, Anto JM, Bel EH, Boulet LP, et al. Severe asthma in adults: what are the important questions? *J Allergy Clin Immunol*. 2007 Jun;119(6):1337-48.
 90. Bel EH, Sousa A, Fleming L, Bush A, Chung F, Versnel J, et al. Diagnosis of severe refractory asthma:an international consensus statement from the Innovative Medicine Initiative (IMI). *Thorax*. 2011;66:910–917.
 91. Wener RL, Bel EH. Severe refractory asthma:an update. *Eur Respir Rev*. 2013;22:196–201.
 92. Chung KF. Asthma phenotyping:a necessity for improved therapeutic precision and new targeted therapies. *J Intern Med*. 2016;279:192–204.
 93. Syabbalo N. Neutrophilic asthma: a complex phenotype of severe asthma. *J Lung*

- Pulm Respir Res. 2020;7(1):18–24.
94. Wenzel SE, Schwartz LB, Langmack EL, J L Halliday, J B Trudeau, R L Gibbs, et al. Evidence that severe asthma can be divided pathologically into two inflammatory subtypes with distinct physiologic and clinical characteristics. *Am J Respir Crit Care Med*. 1999;160:1001–1008.
 95. Simpson JL, Scott R, Boyle MJ, BOYLE MJ, Peter G. Inflammatory subtypes in asthma: assessment and identification using induced sputum. *Respirology*. 2006;11(1):54–61.
 96. Wenzel SE. Asthma phenotypes: the evolution from clinical to molecular approach. *Nat Med*. 2012;18(5):716–725.
 97. Chung K, Godard P, Adelroth E, Ayres J, Barnes N, Barnes P, et al. Difficult/therapy-resistant asthma: the need for integrated approach to define clinical phenotypes, evaluate risk factors, understand pathophysiology and find novel therapies. *Eur Respir J*. 1999;13(5):1198–1208
 98. Pavord ID. Eosinophilic phenotypes of airway disease. *Ann Am Thorac Soc*. 2013;10(Suppl):S143–S149.
 99. Chung KF. Neutrophilic asthma: a distinct target for treatment. *Lancet Respir Med*. 2016;(10):765–767.
 100. Nair P, Aziz-Ur-Rehman A, Radford K. Therapeutic implication of ‘neutrophilic asthma’. *Curr Opin Pulm Med*. 2005;21(1):33–38.
 101. Anderson GP. Endotyping asthma: new insights into key pathogenic mechanism in a heterogenous disease. *Lancet*. 2008;372:1107–1119.
 102. Buhl R, Humbert M, Bjermer L, Chanez P, Heaney LG, Pavord I, et al. Severe eosinophilic asthma: a roadmap to consensus. *Eur Respir J*. 2017;49(5):1700634.
 103. Miranda C, Busacker A, Balzar S, Westcott JY, Trudeau JB, Sun Y, Conrad dj, et al. Distinguishing severe asthma phenotypes: role of onset and eosinophilic inflammation. *J Allergy Clin Immunol*. 2004;113:101–108.
 104. Haldar P, Pavord ID, Shaw DE, Berry MA, Thomas M, Brightling CE, Wardlaw AJ, Green RH. Cluster analysis and clinical asthma phenotypes. *Am J Respir Crit Care Med*. 2008 Aug 1;178(3):218–224.
 105. Groot JC, Ten Brinke A, Bel EHD. Management of patients with eosinophilic asthma: a new era begins. *ERJ Open Res*. 2015;1(1):00024– 2015.
 106. Ray A, Kolls JK. Neutrophilic inflammation in asthma is associated with disease severity. *Trends Immunol*. 2017;38(12):948–954.
 107. Moore WC, Hasle A, Xingnan Li, Huashi Li, William W Busse, Nizar N Jarjour, et al. Sputum neutrophil counts are associated with more severe asthma phenotypes using cluster analysis. *J Allergy Clin Immunol*. 2014;133(6):1557–1563.
 108. Shaw DE, Berry MA, Hargadon B, McKenna S, Shelley MJ, Green RH, et al. Association between neutrophilic airway inflammation and airflow limitation in adults with asthma. *Chest*. 2007;132(6):1871–1875.
 109. Little SA, Macleod KJ, Chalmers GW, Bousquet J, Jeffery PK, Busse WW, et al. Association of forced expired volume with disease duration and sputum neutrophils in chronic asthma. *Am J Med*. 2002;112:446–452.
 110. National Asthma Education and Prevention Program. Expert Panel Report 3 (EPR 3):Guidelines for the Diagnosis and Management of Asthma – A Summary Report 2007. *J Allergy Clin Immunol*. 2007;120(5 Suppl):S94–S138
 111. Fahy JV, Kim KW, Liu J, Boushey HA. Prominent neutrophil inflammation in sputum from subjects with asthma exacerbations. *J Allergy Clin Immunol*.

- 1995;95:843–852.
112. Jayaram L, Pizzichini MM, Cook RJ, Haldar P, Brightling CE, Hargadon B, et al. Determining asthma treatment by monitoring sputum cell counts: effect on exacerbations. *Eur Respir J*. 2006;27(3):483–494.
113. Haldar P, Pavord ID. Noneosinophilic asthma: A distinct clinical and pathologic phenotype. *J Allergy Clin Immunol*. 2007;119(5):1043–1052.
114. Martin RJ, Cicutto LC, Smith HR, Ballard RD, Szefer SJ. Airway inflammation in nocturnal asthma. *Am Rev Respir Dis* 1991;143(2):351–357.
115. Syabbalo NC. Chronobiology and chronopathophysiology of nocturnal asthma. *Int J Clin Pract*. 1997;51 (7), 455–462.
116. Proceeding of the ATS workshop on refractory asthma: current understanding, recommendations, and unanswered questions. American Thoracic Society. *Am J Respir Crit Care Med*. 2000;162(6):2341–2351.
117. Wener RL, Bel EH. Severe refractory asthma: an update. *Eur Respir Rev*. 2013;22:196–201.
118. Siroux V, González JR, Bouzigon E, Curjuric I, Boudier A, Imboden M, et al. Genetic heterogeneity of asthma phenotypes identified by a clustering approach. *Eur Respir J*. 2014;43:439–452
119. Sutherland ER, Goleva E, King TS, Lehman E, Stevens AD, Jackson LP, et al. Cluster analysis of obesity and asthma phenotypes. *PLoS One*. 2012;7(5):e36631.
120. Pavord ID, Brightling CE, Woltmann G, Wardlaw AJ. Non-eosinophilic asthma corticosteroid unresponsive asthma. *Lancet*. 1999;353:2213–2214.
121. Green RH, Brightling CE, Wolmann G, Parker D, Wardlaw AJ, Pavord ID. Analysis of induced sputum in adults with asthma identification of a subgroup with isolated sputum neutrophilia and poor response to inhaled corticosteroids. *Thorax*. 2002;57(10):875–879.
122. Lambrecht BN, Hammad H. The immunology of asthma. *Nat Immunol*. 2015;16(1):45–56.
123. Moore WC, Meyer DA, Wenzel SE, Teague WG, Li H, Li X, D'Agostino R Jr, et al. Identification of asthma phenotypes using cluster analysis in the Severe Asthma Research Program. *Am J Respir Crit Care Med*. 2010;181(4):315–323.
124. Fahy JV. Type 2 inflammation in asthma – present in most, absent in many. *Nat Rev Immunol*. 2015;15(1):57–65.
125. Woodruff PG, Khashayar R, Lazarus SC, Janson S, Avila P, Boushey HA, et al. Relationship between airway inflammation, hyperresponsiveness, and obstructive asthma. *J Allergy Clin Immunol*. 2001;108:753–758.
126. Kuo CS, Pavlidis S, Loza M, Baribaud F, Rowe A, Pandis I, et al. T-helper cell type 2 (Th2) and non- Th2 molecular phenotypes of asthma using sputum transcriptomics in U-BIPRED. *Eur Respir J*. 2017;49(2):1602135.
127. Cosmi L, Liotta F, Maggi E, Romagnani S, Annunziato F. Th17 cells: new players in asthma pathogenesis. *Allergy*. 2011;66(8):989–998.
128. Pavord ID, Korn S, Howarth P, Bleecker ER, Buhl R, Keene ON, et al. Mepolizumab for severe asthma (DREAM): a multicenter, double-blind, placebo-controlled trial. *Lancet*. 2012;380:651–659.
129. Kolls J, Lindén A. Interleukin-17A family members and inflammation. *Immunity*. 2004;21(4):467–476.
130. Lindén A, Laun M, Anderson GK. Neutrophils, interleukin-17A and lung disease. *Eur Respir J*. 2005;25(1):159–172.
131. Brusselle GG, Provoost S, Bracke RR, Kuchmiy A, Lamkanfi M. Inflammasomes

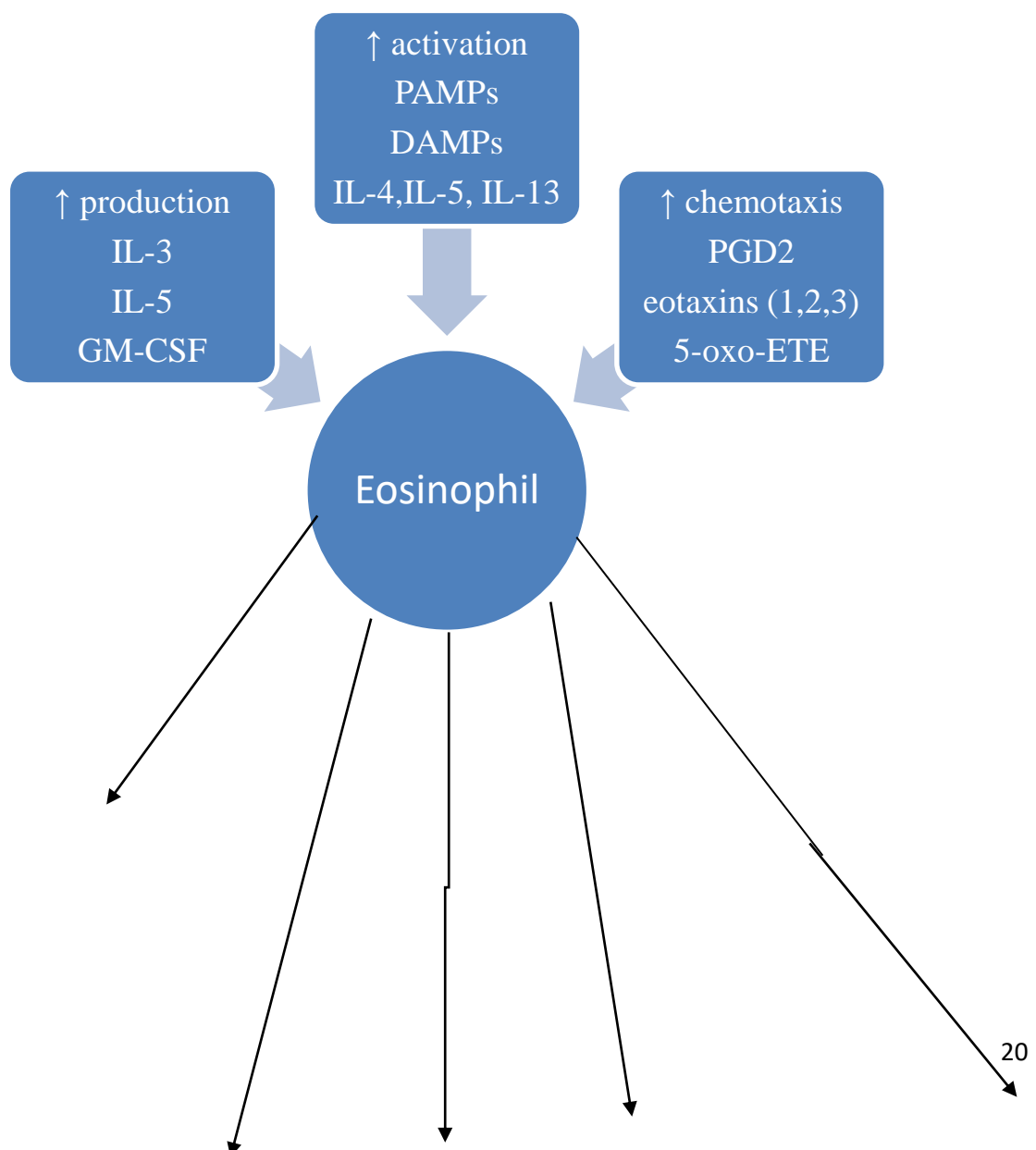
- in respiratory disease:from bench to bedside. *Chest*. 2014;145:1121–1133.
132. Guilleminaut L, Oukseil H, Belleguic C, Yannick Le Guen, Patrick Germaud, Emilie Desfleurs, et al. Personalized medicine in asthma: from curative to preventive medicine. *Eur Respir J*. 2017;2017;26(143):160010.
 133. Ray A, Kolls JK. Neutrophilic Inflammation in Asthma and Association with Disease Severity. *Trends Immunol*. 2017;38(12):942-954.
 134. Raundhal M, Morse C, Khare A, Oriss TB, Milosevic J, Trudeau J, et al. High IFN- γ and low SLPI mark severe asthma in mice and humans. *J Clin Invest*. 2015 Aug 3;125(8):3037-50.
 135. Ray A, Raundhal M, Oriss TB, Ray P, Wenzel SE. Current concepts of severe asthma. *J Clin Invest*. 2016; 126(7):2394–403.
 136. Bullens DM, Truyen E, Coteur L, Dilissen E, Hellings PW, Dupont LJ, Ceuppens JL. IL-17 mRNA in sputum of asthmatic patients: linking T cell driven inflammation and granulocytic influx? *Respir Res*. 2006; 7:135.
 137. McKinley L, Alcorn JF, Peterson A, Dupont RB, Kapadia S, Logar A, et al. TH17 cells mediate steroid-resistant airway inflammation and airway hyperresponsiveness in mice. *J Immunol*. 2008 Sep 15;181(6):4089-97.
 138. Kim HY, Lee HJ, Chang YJ, Pichavant M, Shore SA, Fitzgerald KA, Iwakura Y, et al. Interleukin-17-producing innate lymphoid cells and the NLRP3 inflammasome facilitate obesity-associated airway hyperreactivity. *Nat Med*. 2014; 20(1):54–61. [PubMed: 24336249]
 139. Chen K, Eddens T, Trevejo-Nunez G, Way EE, Elsegeiny W, Ricks DM, Garg AV, et al. IL-17 Receptor Signaling in the Lung Epithelium Is Required for Mucosal Chemokine Gradients and Pulmonary Host Defense against *K. pneumoniae*. *Cell Host Microbe*. 2016; 20(5):596–605.
 140. Dagvadorj J, Shimada K, Chen S, Jones HD, Tumurkhuu G, Zhang W, et al. Lipopolysaccharide Induces Alveolar Macrophage Necrosis via CD14 and the P2X7 Receptor Leading to Interleukin-1 α Release. *Immunity*. 2015; 42(4):640–53.
 141. Wei-Xu H, Wen-yun Z, Xi-ling Z, Zhu W, Li W, Xiao W, et al. Anti-Interleukin-1 Beta/Tumor Necrosis Factor-Alpha IgY Antibodies Reduce Pathological Allergic Responses in Guinea Pigs with Allergic Rhinitis. *Mediators Inflamm*. 2016; 2016:3128182.
 142. Cai S, Batra S, Del Piero F, Jeyaseelan S. NLRP12 modulates host defense through IL-17A-CXCL1 axis. *Mucosal Immunol*. 2016; 9(2):503–14.
 143. Hernandez ML, Mills K, Almond M, Todoric K, Aleman MM, Zhang H, et al. IL-1 receptor antagonist reduces endotoxin-induced airway inflammation in healthy volunteers. *J Allergy Clin Immunol*. 2015; 135(2):379–85.
 144. Hubeau C, Kubera JE, Masek-Hammerman K, Williams CM. Interleukin-6 neutralization alleviates pulmonary inflammation in mice exposed to cigarette smoke and poly(I:C). *Clin Sci (Lond)*. 2013; 125(10):483–93.
 145. Li HD, Zhang QX, Mao Z, Xu XJ, Li NY, Zhang H. Exogenous interleukin-10 attenuates hyperoxia-induced acute lung injury in mice. *Exp Physiol*. 2015; 100(3):331–40.
 146. Hsu CY, Liu HE, Sheu FY, Leu SJ, Chiang BL, Hsiao G, Lee YL. Synergistic therapeutic effects of combined adenovirus-mediated interleukin-10 and interleukin-12 gene therapy on airway inflammation in asthmatic mice. *J Gene Med*. 2010; 12(1):11–21.
 147. Wakashin H, Hirose K, Maezawa Y, Kagami S, Suto A, Watanabe N, et al. IL-23

- and Th17 cells enhance Th2-cell-mediated eosinophilic airway inflammation in mice. *Am J Respir Crit Care Med*. 2008; 178(10):1023–32.
148. Dubin PJ, Martz A, Eisenstatt JR, Fox MD, Logar A, Kolls JK. Interleukin-23-mediated inflammation in *Pseudomonas aeruginosa* pulmonary infection. *Infect Immun*. 2012; 80(1):398–409.
 149. Nguyen TH, Maltby S, Simpson JL, Eysers F, Baines KJ, Gibson PG, et al. TNF-alpha and Macrophages Are Critical for Respiratory Syncytial Virus-Induced Exacerbations in a Mouse Model of Allergic Airways Disease. *J Immunol*. 2016; 196(9): 3547–58.
 150. Fei M, Bhatia S, Oriss TB, Yarlagadda M, Khare A, Akira S, et al. TNF-alpha from inflammatory dendritic cells (DCs) regulates lung IL-17A/IL-5 levels and neutrophilia versus eosinophilia during persistent fungal infection. *Proc Natl Acad Sci U S A*. 2011; 108(13):5360–5.
 151. Hicks A, Goodnow Jr R, Cavallo G, Tannu SA, Ventre JD, Lavelle D, et al. Effects of LTB4 receptor antagonism on pulmonary inflammation in rodents and non-human primates. *Prostaglandins Other Lipid Mediat*. 2010; 92(1-4):33–43.
 152. Karp CL, Flick LM, Park KW, Softic S, Greer TM, Keledjian R, et al. Defective lipoxin-mediated anti-inflammatory activity in the cystic fibrosis airway. *Nat Immunol*. 2004; 5(4):388–92.
 153. Serhan CN, Chiang N, Dalli J, Levy BD. Lipid mediators in the resolution of inflammation. *Cold Spring Harb Perspect Biol*. 2014; 7(2):a016311.
 154. Haworth O, Cernadas M, Yang R, Serhan CN, Levy BD. Resolvin E1 regulates interleukin 23, interferon-gamma and lipoxin A4 to promote the resolution of allergic airway inflammation. *Nat Immunol*. 2008; 9(8):873–9.
 155. Jandl K, Stacher E, Bálint Z, Sturm EM, Maric J, Peinhaupt M, et al. Activated prostaglandin D2 receptors on macrophages enhance neutrophil recruitment into the lung. *J Allergy Clin Immunol*. 2016; 137(3):833–43.
 156. Lombard R, Doz E, Carreras F, Eparaud M, Vern YL, Gatel DB, et al. IL-17RA in Non-Hematopoietic Cells Controls CXCL-1 and 5 Critical to Recruit Neutrophils to the Lung of Mycobacteria-Infected Mice during the Adaptive Immune Response. *PLoS One*. 2016; 11(2):e0149455.
 157. Trevejo-Nunez G, Chen K, Dufour JP, Bagby GJ, Horne WT, Nelson S, et al. Ethanol impairs mucosal immunity against *Streptococcus pneumoniae* infection by disrupting interleukin 17 gene expression. *Infect Immun*. 2015; 83(5):2082–8.
 158. Prause O, Laan M, Lötvall J, Lindén A. Pharmacological modulation of interleukin-17-induced GCP-2-, GRO-alpha- and interleukin-8 release in human bronchial epithelial cells. *Eur J Pharmacol*. 2003; 462(1-3):193–8.
 159. Rohde G, Message SD, Haas JJ, Keadze T, Parker H, Laza-Stanca V, et al. CXC chemokines and antimicrobial peptides in rhinovirus-induced experimental asthma exacerbations. *Clin Exp Allergy*. 2014; 44(7):930–9.
 160. Staab EB, Sanderson SD, Wells SM, Poole JA. Treatment with the C5a receptor/CD88 antagonist PMX205 reduces inflammation in a murine model of allergic asthma. *Int Immunopharmacol*. 2014; 21(2):293–300.
 161. Sun L, Guo RF, Gao H, Sarma JV, Zetoune FS, Ward PA. Attenuation of IgG immune complex-induced acute lung injury by silencing C5aR in lung epithelial cells. *FASEB J*. 2009; 23(11):3808–18.
 162. Tjabringa GS, Ninaber DK, Drijfhout JW, Rabe KF, Hiemstra PS. Human cathelicidin LL-37 is a chemoattractant for eosinophils and neutrophils that acts via formyl-peptide receptors. *Int Arch Allergy Immunol*. 2006; 140(2):103–12.

163. Myou S, Leff AR, Myo S, Boetticher E, Tong J, Meliton AY. Blockade of inflammation and airway hyperresponsiveness in immune-sensitized mice by dominant-negative phosphoinositide 3-kinase-TAT. *J Exp Med.* 2003; 198(10):1573–82.
164. Kunkel SL, Standiford T, Kasahara K, Strieter RM. Interleukin-8 (IL-8): the major neutrophil chemotactic factor in the lung. *Exp Lung Res.* 1991; 17(1):17–23.
165. Teran LM, Johnston SL, Schröder JM, Church MK, Holgate ST. Role of nasal interleukin-8 in neutrophil recruitment and activation in children with virus-induced asthma. *Am J Respir Crit Care Med.* 1997; 155(4):1362–6.
166. Gibson PG, Simpson JL, Saltos N. Heterogeneity of airway inflammation in persistent asthma: evidence of neutrophilic inflammation and increased sputum interleukin-8. *Chest.* 2001; 119(5):1329–36.
167. Gounni AS, Lamkhoui B, Koussih L, Ra C, Renzi PM, Hamid Q. Human neutrophils express the high-affinity receptor for immunoglobulin E (Fc epsilon RI): role in asthma. *FASEB J.* 2001; 15(6):940–9.
168. Nauseef WM, Borregaard N. Neutrophils at work. *Nat Immunol.* 2014; 15(7):602–11.
169. Soehnlein O, Weber C, Lindbom L. Neutrophil granule proteins tune monocytic cell function. *Trends Immunol.* 2009; 30(11):538–46.
170. Nadel JA. Role of neutrophil elastase in hypersecretion during COPD exacerbations, and proposed therapies. *Chest.* 2000; 117(5 Suppl 2):386S–9S.
171. Anticevich SZ, Hughes JM, Black JL, Armour CL. Induction of hyperresponsiveness in human airway tissue by neutrophils-- mechanism of action. *Clin Exp Allergy.* 1996; 26(5):549–56.
172. Stănescu D, Sanna A, Veriter C, Kostianev S, Calcagni PG, Fabbri LM, et al. Airways obstruction, chronic expectoration, and rapid decline of FEV1 in smokers are associated with increased levels of sputum neutrophils. *Thorax.* 1996; 51(3):267–71.
173. Lezmi G, Gosset P, Deschildre A, Abou-Taam R, Mahut B, Beydon N, et al. Airway Remodeling in Preschool Children with Severe Recurrent Wheeze. *Am J Respir Crit Care Med.* 2015; 192(2):164–71.
174. Grainge CL, Lau LCK, Ward JA, Dulay V, Lahiff G, Wilson S, et al. Effect of bronchoconstriction on airway remodeling in asthma. *N Engl J Med.* 2011; 364(21):2006–15.
175. Cox G. Glucocorticoid treatment inhibits apoptosis in human neutrophils. Separation of survival and activation outcomes. *J Immunol.* 1995; 154(9):4719–25.
176. Nguyen LT, Lim S, Oates T, Chung KF. Increase in airway neutrophils after oral but not inhaled corticosteroid therapy in mild asthma. *Respir Med.* 2005; 99(2):200–7.
177. Louis R, Lau LC, Bron AO, Roldaan AC, Radermecker M, Djukanović R. The relationship between airways inflammation and asthma severity. *Am J Respir Crit Care Med.* 2000; 161(1):9–16.
178. Fukakusa M, Bergeron C, Tulic MK, Fiset PO, Al Dewachi O, Laviolette M, et al. Oral corticosteroids decrease eosinophil and CC chemokine expression but increase neutrophil, IL-8, and IFN-gamma-inducible protein 10 expression in asthmatic airway mucosa. *J Allergy Clin Immunol.* 2005; 115(2):280–6.
179. Dubyak GR, el-Moatassim C. Signal transduction via P2-purinergic receptors for

- extracellular ATP and other nucleotides. *Am J Physiol.* 1993; 265(3 Pt 1):C577–606.
- 180.Uddin M, Nong G, Ward J, Seumois G, Prince LR, Wilson SJ, et al. Prosurvival activity for airway neutrophils in severe asthma. *Thorax.* 2010; 65(8): 684–9.
- 181.Vaughan KR, Stokes L, Prince LR, Marriott HM, Meis S, Kassack MU, et al. Inhibition of neutrophil apoptosis by ATP is mediated by the P2Y11 receptor. *J Immunol.* 2007; 179(12):8544–53.
- 182.Lee E, Lindo T, Jackson N, Meng-Choong L, Reynolds P, Hill A, et al. Reversal of human neutrophil survival by leukotriene B(4) receptor blockade and 5-lipoxygenase and 5-lipoxygenase activating protein inhibitors. *Am J Respir Crit Care Med.* 1999; 160(6):2079–85.
- 183.Thomas CJ, Schroder K. Pattern recognition receptor function in neutrophils. *Trends Immunol.* 2013; 34(7):317–28.
- 184.Kritas SK, Cerulli ASG, Speziali A, Antinolfi P, Pantalone A, Rosati M, et al. Asthma and mast cell biology. *Eur J Inflamm* 2014;12(2):261-265.
- 185.Weller CL, Collington SJ, Williams T, Lamb JR. Mast cells in health and disease. *Clin Sci (Lond)* 2011; 120(11):473-84.
- 186.Kandere-Grzybowska K, Letourneau R, Kempuraj D, Donelan J, Poplawski S, Boucher W, et al. IL-1 induces vesicular secretion of IL-6 without degranulation from human mast cells. *J Immunol* 2003; 171(9):4830-6.
- 187.Conti P, Pang X, Boucher W, Letourneau R, Reale M, Barbacane RC, et al. Impact of Rantes and MCP-1 chemokines on in vivo basophilic cell recruitment in rat skin injection model and their role in modifying the protein and mRNA levels for histidine decarboxylase. *Blood* 1997; 89(11):4120-7.
- 188.Irani AM, Schwartz LB. Human mast cell heterogeneity. *Allergy Proc* 1994; 15(6):303-8.
- 189.Hamid Q, Tulic M. Immunobiology of asthma. *Annu Rev Physiol* 2009; 71:489-507.
- 190.Hsu, F.; Boyce, JA. Biology of mast cells and their mediators. In: Adkinson, N.; Bochner, BS.; Busse, WW.; Holgate, ST.; Lemanske, RF.; Simons, FER., editors. *Middleton's Allergy: Principles and Practice*. 7th ed. Mosby Elsevier; 2009.
- 191.Metz, M.; Brockow, K.; Metcalfe, DD.; Galli, SJ. Mast cells, basophils, and mastocytosis. 3rd ed. Rich, R.; Fleisher, TA.; Shearer, WT.; Schroeder, HW.; Frew, AJ.; Weyand, CM., editors. Mosby Elsevier; 2008.
- 192.Metcalfe DD. Mast cells and mastocytosis. *Blood* 2008;112:946–56.
- 193.Theoharides TC, Conti P. Mast cells: the Jekyll and Hyde of tumor growth. *Trends Immunol* 2004; 25(5):235-41.
- 194.Cho WS, Chae C. Expression of inflammatory cytokines (TNF- α , IL-1, IL-6 and IL-8) in colon of pigs naturally infected with *Salmonella typhimurium* and *S. choleraesuis*. *J Vet Med A Physiol Pathol Clin Med* 2003; 50(10):484-7.
- 195.Theoharides TC, Enakua S, Sismanopoulos N, Asadi S, Papadimas EC, Angelidou A, Alysandratos KD. Contribution of stress to asthma worsening through mast cell activation. *Ann Allergy Asthma Immunol* 2012; 109(1):14-9.
- 196.Matsuzawa S, Sakashita K, Kinoshita T, Ito S, Yamashita T, Koike K. IL-9 enhances the growth of human mast cell progenitors under stimulation with stem cell factor. *J Immunol* 2003; 170(7):3461-7.
- 197.Yoshimoto T, Tsutsui H, Tominaga K, Hoshino K, Okamura H, Akira S, et al. IL-18, although antiallergic when administered with IL-12, stimulates IL-4 and histamine release by basophils. *Proc Natl Acad Sci U S A* 1999; 96(24):13962-6.

198. Fux M, Pecaric-Petkovic T, Odermatt A, Hausmann OV, Lorentz A, Bischoff SC, et al. IL-33 is a mediator rather than a trigger of the acute allergic response in humans. *Allergy* 2014; 69(2):216-22.
199. Mrabet-Dahbi S, Metz M, Dudeck A, Zuberbier T, Maurer M. Murine mast cells secrete a unique profile of cytokines and prostaglandins in response to distinct TLR2 ligands. *Exp Dermatol* 2009; 18(5):437-44.
200. Sismanopoulos N, Delivanis DA, Mavrommati D, Hatziagelaki E, Conti P, Theoharides TC. Do mast cells link obesity and asthma? *Allergy* 2013; 68(1):8-15.
201. Bergqvist A, Andersson CK, Mori M, Walls AF, Bjermer L, Erjefalt JS. Alveolar T-helper type-2 immunity in atopic asthma is associated with poor clinical control. *Clin Sci (Lond)*. 2015;128(1):47-56. doi:10.1042/CS20140309.
202. Kiyokawa H, Matsumoto H, Nakaji H, Niimi A, Ito I, Ono K, et al. Centrilobular opacities in the asthmatic lung successfully treated with inhaled ciclesonide and tiotropium: with assessment of alveolar nitric oxide levels. *Allergol Int*. 2011;60(3):381-5. doi:10.2332/allergolint.10-CR-0251



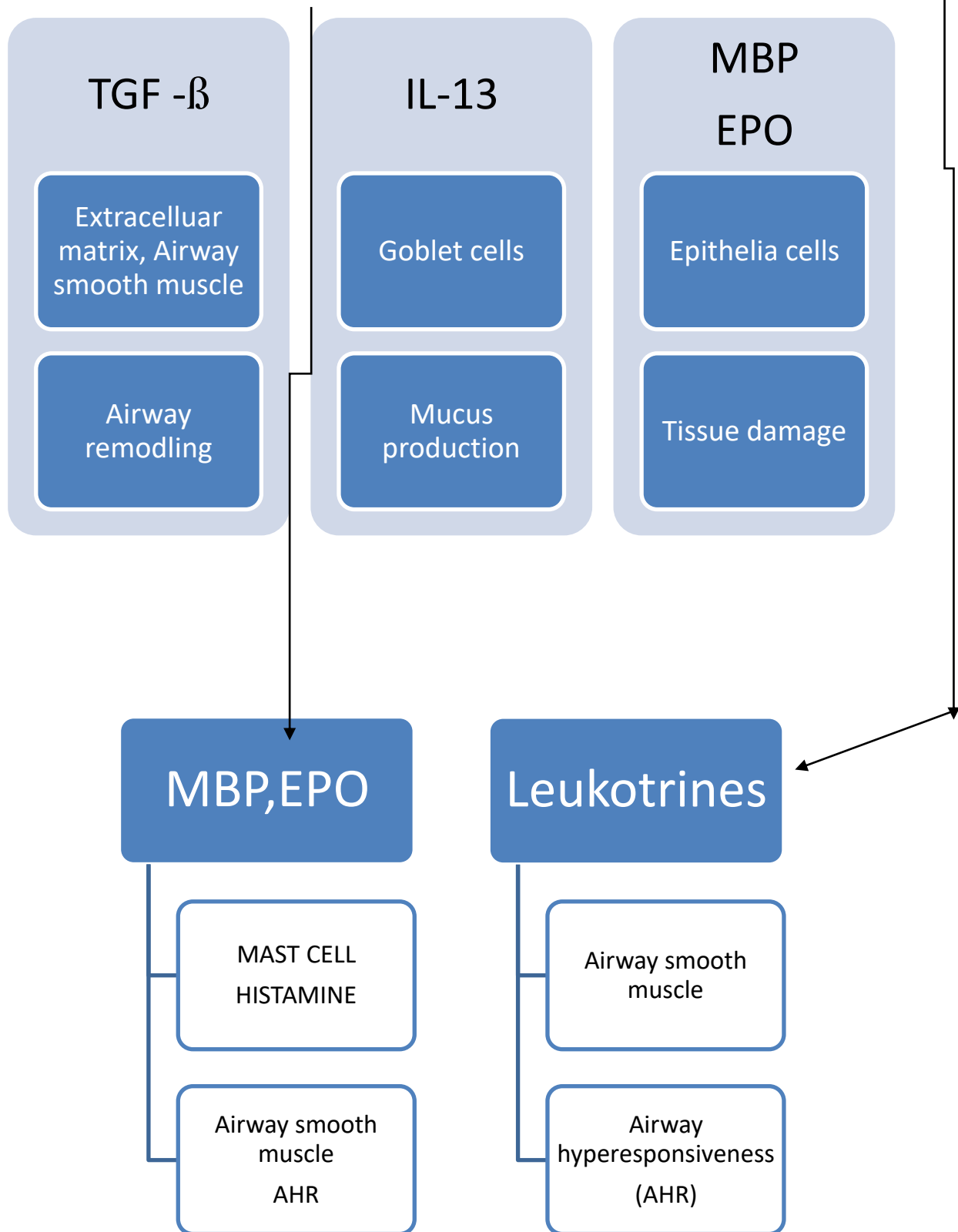


Fig.1. Eosinophils role in asthma pathogenesis [15]